

## Phenyl and Phenylethyl Glycosides from *Picrorhiza scrophulariiflora*

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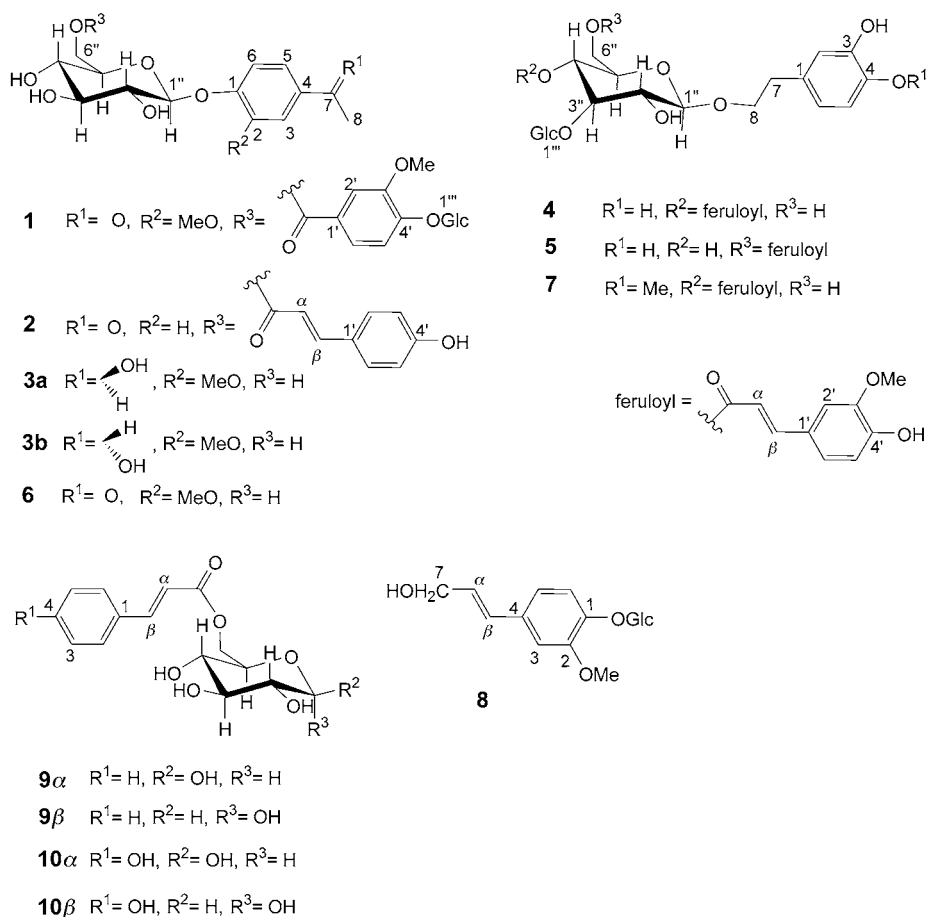
Three new phenyl glycosides, scrophenoside A (**1**), B (**2**), and C (**3**), and two new phenylethyl glycosides, scroside D (**4**) and scroside E (**5**), were isolated from the stem of *Picrorhiza scrophulariiflora* PENNELL (Scrophulariaceae), besides five known compounds. On the basis of spectroscopic evidence, the structures of the new compounds were elucidated as 4-acetyl-2-methoxyphenyl 6-*O*-[4-( $\beta$ -D-glucopyranosyloxy)vanilloyl]- $\beta$ -D-glucopyranoside (**1**), 4-acetylphenyl 6-*O*-[(*E*)-*p*-coumaroyl]- $\beta$ -D-glucopyranoside (**2**), 4-[(1*R*)- and (1*S*)-1-hydroxyethyl]-2-methoxyphenyl  $\beta$ -D-glucopyranoside (**3a** and **3b**, resp.), 2-(3,4-dihydroxyphenyl)ethyl *O*- $\beta$ -D-glucopyranosyl-(1  $\rightarrow$  3)-4-*O*-[(*E*)-feruloyl]- $\beta$ -D-glucopyranoside (**4**), and 2-(3,4-dihydroxyphenyl)ethyl *O*- $\beta$ -D-glucopyranosyl-(1  $\rightarrow$  3)-6-*O*-[(*E*)-feruloyl]- $\beta$ -D-glucopyranoside (**5**).

**Introduction.** – *Picrorhiza scrophulariiflora* PENNELL and *Picrorhiza kurrooa* ROYLE ex BENTH (Scrophulariaceae) are extensively used in traditional medicine of China, Nepal, and India for the treatment of various immune-system-related diseases. For instance, both plants are used in asthma, jaundice, and arthritis [1]. Previous phytochemical investigation of this plant revealed the presence of iridoid glycosides, cucurbitacins, cucurbitacin glycosides, and phenyl glycosides [2–5]. In this paper, we describe the isolation and identification of three new phenyl glycosides and two new phenylethyl glycosides as well as of five known compounds, androsin (**6**) [4][6], hemiphroside A (**7**) [7], coniferin (**8**) [8], 6-*O*-cinnamoyl-D-glucopyranose (**9**) [9], and 6-*O*-(*p*-coumaroyl)-D-glucopyranose (**10**) [10], which were isolated from the EtOH extract of the stems of *Picrorhiza scrophulariiflora*.

**Results and Discussion.** – Scrophenoside A (**1**) was obtained as a white amorphous powder. Its molecular formula was determined as C<sub>29</sub>H<sub>36</sub>O<sub>16</sub> by HR-FAB-MS ( $[M - H]^-$  at  $m/z$  639.1926, calc. 639.1925). The IR absorption indicated the presence of a carbonyl group, an ester group (1704, 1678 cm<sup>-1</sup>), and an aromatic ring (1593, 1512, 1465, 1418 cm<sup>-1</sup>). The structure of **1** (Fig. 1) was established by <sup>1</sup>H- and <sup>13</sup>C-NMR data, DEPT, HSQC, HMBC, and tandem MS experiments as 4-acetyl-2-methoxyphenyl 6-*O*-[4-( $\beta$ -D-glucopyranosyloxy)vanilloyl]- $\beta$ -D-glucopyranoside<sup>1)</sup>.

The <sup>1</sup>H-NMR spectrum of **1** showed the presence of two 1,2,4-trisubstituted benzene rings, which was confirmed by two *ABX* systems at  $\delta$  7.14–7.52 for six aromatic protons ( $\delta$  7.44 (*d*, *J* = 1.6 Hz), 7.30 (*dd*, *J* = 1.6, 8.5 Hz), 7.14 (*d*, *J* = 8.5 Hz);  $\delta$  7.45 (*d*, *J* = 1.7 Hz), 7.52 (*dd*, *J* = 1.7, 8.5 Hz), 7.18 (*d*, *J* = 8.5 Hz)). A comparison of the <sup>1</sup>H- and <sup>13</sup>C-NMR data with those of **6** and vanillic acid (= 4-hydroxy-3-methoxybenzoic acid) suggested that the two 1,2,4-trisubstituted benzene rings were part of an androsin and a vanilloyl moiety. In addition, two

<sup>1)</sup> Arbitrary numbering; for systematic names, see *Exper. Part*

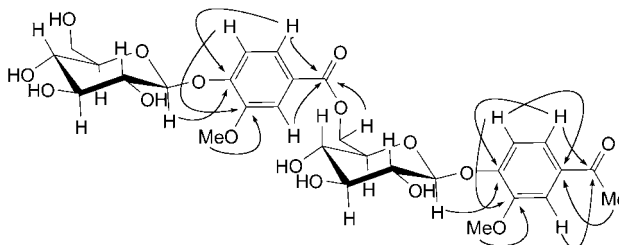
Fig. 1. Structure of compounds **1–10**.)

anomeric protons were observed at  $\delta$  5.13 ( $d, J = 7.2$  Hz) and 5.04 ( $d, J = 7.2$  Hz) indicating the diglycosidic structure of **1**, which was also confirmed by the  $^{13}\text{C}$ -NMR spectrum of **1** ( $\delta$  100.16 and 99.7; Table). To determine their connectivities, HMBC and tandem ESI-MS experiments were employed. As shown in Fig. 2, the long-range correlations of the two anomeric protons ( $\delta$  5.13 and 5.04) with C(1) ( $\delta$  150.93) and C(4') ( $\delta$  151.42), respectively, indicated that two glucose units were connected with C(1) and C(4')<sup>1</sup>. Likewise, the correlations of  $\text{CH}_2(6'')$  with C=O ( $\delta$  165.76) enabled us to assign the position of the vanilloyl group to C(6'') of glucose. The structure of **1** was further confirmed by an ESI-MS<sup>n</sup> experiment. MS<sup>1</sup> gave the pseudomolecular ion at  $m/z$  675 ( $[M + \text{Cl}]^-$ ), MS<sup>2</sup> the fragment ion at  $m/z$  477 ( $[M - \text{H} - 162]^-$ ), and MS<sup>3</sup> the fragment ions at  $m/z$  311 ( $[M - \text{H} - 162 - 166]^-$ ) and 165 ( $[M - \text{H} - 162 - 166 - 146]^-$ ), corresponding to the sequential loss of a glucose unit, and the vanillic acid and glucose moieties.

Scrophenoside B (**2**) was isolated as a white amorphous powder. Its molecular formula was determined as  $\text{C}_{23}\text{H}_{24}\text{O}_9$  by HR-FAB-MS ( $[M - \text{H}]^-$  at  $m/z$  443.1355, calc. 443.1342). The IR absorption indicated the presence of a carbonyl and an ester group ( $1693, 1659\text{ cm}^{-1}$ ), a C=C bond ( $1610\text{ cm}^{-1}$ ), and an aromatic ring ( $1582, 1516, 1449\text{ cm}^{-1}$ ). Further investigation of 1D- and 2D-NMR data resulted in the

Table 1.  $^{13}\text{C}$ -NMR Data ( $(\text{D}_6)$ DMSO) for **1–6** and **7**.  $\delta$  in ppm $^1$ .

	<b>1</b>	<b>2</b>	<b>3a</b>	<b>3b</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>
C(1)	150.93	161.43	145.76	145.76	129.11	129.87	151.02	130.94
C(2)	149.28 <sup>a)</sup>	116.49 <sup>a)</sup>	149.35	149.35	116.28	116.96	149.49	116.19
C(3)	111.66	130.86	110.47	110.52	144.96	145.62	111.90	145.98
C(4)	131.51	131.49	141.98	141.98	143.51	144.17	131.74	144.72
C(5)	122.87	130.86	117.88	117.88	115.40	116.08	123.11	112.17
C(6)	114.75	116.49 <sup>a)</sup>	115.76	115.76	119.54	120.15	114.82	119.30
C(7)	197.13	197.00	68.47	68.51	34.98	35.80	197.36	34.86
C(8)	27.08	26.89	26.69	26.69	70.12	70.80	27.19	69.90
MeO	56.25 <sup>b)</sup>	–	56.25	56.25	–	–	56.26	55.50
C(1')	123.62	125.69	100.85	100.91	125.78	126.21	–	125.71
C(2')	113.43	131.00	73.89	73.89	110.98	111.70	–	110.84
C(3')	149.25 <sup>a)</sup>	116.50 <sup>a)</sup>	77.64	77.64	147.84	148.62	–	147.80
C(4')	151.42	160.57	70.34	70.34	149.35	150.04	–	149.08
C(5')	114.97	131.00	77.53	77.53	115.42	116.13	–	115.36
C(6')	123.58	116.50 <sup>a)</sup>	61.34	61.34	123.02	124.10	–	122.98
C=O	165.76	167.00	–	–	165.84	167.28	–	165.78
C( $\alpha$ )	–	114.69	–	–	114.84	114.88	–	114.63
C( $\beta$ )	–	145.51	–	–	144.76	145.99	–	144.72
MeO	56.41 <sup>b)</sup>	–	–	–	55.59	56.40	–	55.59
C(1'')	100.16	100.12	–	–	102.11	102.86	100.16	102.03
C(2'')	73.64	73.72	–	–	74.57 <sup>a)</sup>	72.77	74.04	74.51
C(3'')	77.30	77.00	–	–	82.91	88.31	77.31	82.85
C(4'')	70.70	70.67	–	–	69.30	69.15	70.71	69.27
C(5'')	74.58	74.53	–	–	74.36 <sup>a)</sup>	73.82	77.83	74.26
C(6'')	64.62	63.94	–	–	60.94	63.87	61.65	60.86
C(1''')	99.70	–	–	–	104.48	104.74	–	104.44
C(2''')	73.74	–	–	–	73.26	74.50	–	73.19
C(3''')	77.73	–	–	–	76.28	76.71	–	76.22
C(4''')	70.14	–	–	–	69.88	70.82	–	69.79
C(5''')	77.51	–	–	–	76.88	77.59	–	76.84
C(6''')	61.19	–	–	–	60.75	61.74	–	60.68

<sup>a)</sup> <sup>b)</sup> Signals in each vertical column are interchangeable.Fig. 2. Key HMBC correlations of **1**

determination of the structure of **2** as 4-acetylphenyl 6-*O*-[(*E*)-*p*-coumaroyl]- $\beta$ -D-glucopyranoside<sup>1)</sup> (*p*-coumaric acid = (2*E*)-3-(4-hydroxyphenyl)-prop-2-enoic acid).

The  $^1\text{H}$ -NMR spectrum of **2** exhibited two *AA'**BB'* systems ( $\delta$  7.84 (*d*, *J* = 8.4 Hz, 2 H), 7.09 (*d*, *J* = 8.4 Hz, 2 H);  $\delta$  7.54 (*d*, *J* = 8.4 Hz, 2 H), 6.79 (*d*, *J* = 8.4 Hz, 2 H), a Me group ( $\delta$  2.37 (*s*)), an (*E*)-double bond ( $\delta$  7.53

( $d, J = 15.9$  Hz, 1 H), 6.38 ( $d, J = 15.9$  Hz, 1 H), and an anomeric proton ( $\delta$  5.06  $d, J = 7.2$  Hz, 1 H). Combined with the  $^{13}\text{C}$ -NMR data (Table), these data suggested that **2** was a phenyl glycoside containing two 1,4-disubstituted benzene rings. In the HMBC spectrum, the anomeric proton H-C(1'') showed correlation with C(1), and the correlation of CH<sub>2</sub>(6'') with C=O ( $\delta$  167.00) was also observed<sup>1</sup>). Thus we concluded that an ester group was connected with C(6'') of the glucose unit. The structure of **2** was further supported by the ESI-MS<sup>n</sup> analysis. MS<sup>1</sup> gave the quasimolecular ion at  $m/z$  443 ( $[M - H]^-$ , MS<sup>2</sup> the fragment ion at  $m/z$  307 ( $[M - H - 136]^-$ ), and MS<sup>3</sup> the fragment ions at  $m/z$  163 and 145.

Scrophenoside C (**3**), an amorphous powder, was isolated as a diastereoisomer mixture **3a/3b**. Its molecular formula was determined as C<sub>15</sub>H<sub>22</sub>O<sub>8</sub> by HR-FAB-MS ( $[M - H]^-$  at  $m/z$  329.1265, calc. 329.1236). On the basis of spectral evidence and chemical transformation, compound **3** was identified as 4-[(1*R*)- and (1*S*)-1-hydroxyethyl]-2-methoxyphenyl  $\beta$ -D-glucopyranoside.

The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data of **3** showed the presence of a glucose, and a 1,2,4-trisubstituted benzene ring. In the  $^1\text{H}$ -NMR spectrum, a Me signal was present at  $\delta$  1.29 ( $d, J = 6.3$  Hz), a CH resonance at  $\delta$  4.63 ( $q, J = 6.3$  Hz), a MeO signal at  $\delta$  3.72 ( $s$ ), and an anomeric proton at  $\delta$  4.82 ( $d, J = 7.2$  Hz, 1 H). In the HMBC spectrum (Fig. 3), the correlations further confirmed the structure of **3**. Furthermore, in the  $^{13}\text{C}$ -NMR spectra (Table), C(7), C(3), and C(1) appeared as split signals, so we concluded that **3** might be a mixture of (7*R*) and (7*S*)-diastereoisomers<sup>1</sup>). To confirm this deduction, we reduced androsin (**6**) with NaBH<sub>4</sub> to a diastereoisomer mixture which was identical to scrophenoside C (**3**) (by  $^{13}\text{C}$ -NMR and TLC).

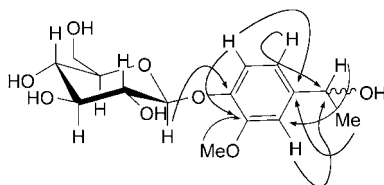
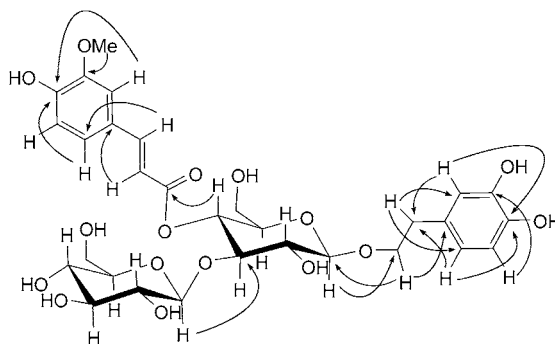


Fig. 3. Key HMBC correlations for **3**

Scroside D (**4**), a yellow amorphous powder, was determined as C<sub>30</sub>H<sub>38</sub>O<sub>16</sub> by HR-FAB-MS ( $[M - H]^-$  at  $m/z$  653.2099, calc. 653.2082). The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of **4** (see Exper. Part and Table) were similar to those of the known compound **7** [7], except for the absence of 1 MeO group at C(4), indicating that **4** might be 2-(3,4-dihydroxyphenyl)ethyl *O*- $\beta$ -D-glucopyranosyl-(1  $\rightarrow$  3)-4-*O*-[(*E*)-feruloyl]- $\beta$ -D-glucopyranoside<sup>1</sup>) (ferulic acid = (2*E*)-3-(4-hydroxy-3-methoxyphenyl)prop-2-enoic acid). The DEPT, HSQC, HMBC, and ESI-MS-MS data confirmed the structure of **4**.

The long-range correlation of C(3') ( $\delta$  147.84) with MeO ( $\delta$  3.80) (Fig. 4) and the fragment ion at  $m/z$  477 ( $[M - 177]^-$ , in the ESI-MS-MS (MS<sup>2</sup>) of **4** indicated the presence of a feruloyl group. Also the correlations of the ester C=O with H-C(4''), of C(3'') ( $\delta$  82.91) with H-C(1'') ( $\delta$  4.37), and of C(8) ( $\delta$  70.12) with H-C(1'') ( $\delta$  4.39) were also observed.

Scroside E (**5**) was isolated as a yellow amorphous powder. Its HR-FAB-MS spectrum ( $[M - H]^-$  at  $m/z$  653.2080, calc. 653.2082) indicated that it had the same molecular formula as **4** (C<sub>30</sub>H<sub>38</sub>O<sub>16</sub>). The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of **5** were similar to those of **4**, and detailed analysis of the HSQC, HMBC, and tandem ESI-MS data as well as the hydrolysis results suggested that **5** was an isomer of **4**; i.e., 2-(3,4-dihydroxyphenyl)ethyl *O*- $\beta$ -D-glucopyranosyl-(1  $\rightarrow$  3)-6-*O*-[(*E*)-feruloyl]- $\beta$ -D-glucopyranoside<sup>1</sup>).

Fig. 4. Key HMBC correlations for **4**

The  $^1\text{H}$ -NMR spectrum of **5** showed a marked upfield shift ( $\delta$  3.34) of  $\text{H}-\text{C}(4'')$  and significant downfield shifts ( $\delta$  4.21 and 4.39) of  $\text{CH}_2(6'')$  for glucose, compared with the corresponding signals of **4** suggesting that the feruloyl moiety should be connected to  $\text{C}(6'')$ . Moreover, the long-range correlations of the ester  $\text{C}=\text{O}$  with  $\text{CH}_2(6'')$  in the HMBC spectrum of **5** further confirmed the position of the feruloyl at  $\text{C}(6'')$  of the glucose unit. On hydrolysis of **5**, only  $\beta$ -D-glucose was obtained (by TLC).

All known compounds were identified by comparing their physical and spectral data with literature values. The main constituent of *P. scrophulariiflora*, androsin (**6**), which was also isolated from *P. kurrooa*, was shown to possess anti-asthma activities [11]. The structure of the new compounds scrophenoside A (**1**), B (**2**), and C (**3**) are similar to androsin; thus they may be the bioactive anti-asthma components in *P. scrophulariiflora*. In addition, phenylethyl glycosides, which were abundant in this plant but had not been found in *P. kurrooa*, seem to be the characteristic constituents in the genus of *P. scrophulariiflora*.

#### Experimental Part

**General.** Column chromatography (CC): *Sephadex LH-20* (Pharmacia), silica gel (200–300 mesh; Qingdao Marine Chemical Group Co.), *Lobar LiChroprep RP-18* (40–63  $\mu\text{m}$ , Merck), *Lobar LiChroprep Si-60* (40–63  $\mu\text{m}$ , Merck), and *MCI-gel CHP-20P* (75–150  $\mu\text{m}$ , Mitsubishi Chemical Co.). Optical rotations: *Perkin-Elmer 341* polarimeter. IR Spectra: *Perkin-Elmer Spectrum-One* FT-IR spectrometer;  $\tilde{\nu}$  in  $\text{cm}^{-1}$ . NMR Spectra: *Bruker AV-600* spectrometer;  $\delta$  in ppm rel. to  $\text{SiMe}_4$  as internal standard,  $J$  in Hz. MS: *VGAutoSpec 3000* spectrometer for HR-FAB and *Finnigan LCQ-Deca* for ESI-MS and tandem spectra.

**Plant Material.** The dried stems of *Picrorhiza scrophulariiflora* were collected from Tibet and identified by Prof. Zhao Zuo-Cheng. A voucher specimen was deposited in the Herbarium of Chengdu Institute of Biology, Chinese Academy of Sciences.

**Extraction and Isolation.** The dried stems of *Picrorhiza scrophulariiflora* (6 kg) were powdered and extracted with EtOH at  $60^\circ$  ( $3 \times 8$  h). After filtration, the solvent was evaporated to give 2 kg of extract. The extract was suspended in  $\text{H}_2\text{O}$  and extracted with  $\text{CHCl}_3$  and AcOEt/MeOH 10:1. The latter extract (50 g) was submitted to CC (silica gel,  $\text{CHCl}_3/\text{MeOH}$  of increasing polarity): *Fractions 1–4*. *Fr. 1* (5 g) was subjected to CC (ODS gel, MeOH/ $\text{H}_2\text{O}$  5:1, 3:1, and 1:1): *Fr. 1.1–1.3*. Every fraction was submitted to CC (*RP-18*): **6** (500 mg), **9** (= **9 $\alpha$ /9 $\beta$** ) (14 mg), and **2** (15 mg), after purification by CC (*Sephadex LH-20* MeOH). *Fr. 2* (25 g) was separated by CC (*MCI-gel* MeOH/ $\text{H}_2\text{O}$  gradient 1:5  $\rightarrow$  8:1): *Fr. 2.1–2.7*. *Fr. 2.2* afforded **3** (10 mg) and **8** (60 mg) after CC (silica gel (*Si-60*), AcOEt/MeOH 15:1). *Fr. 2.4* was submitted CC (*RP-18*): **10** (= **10 $\alpha$ /10 $\beta$** ) (12 mg). *Fr. 2.6* was submitted to CC (silica gel  $\text{CHCl}_3/\text{MeOH}$  8:1): **7** (35 mg). *Fr. 3* (10 g) was separated by CC (silica gel, AcOEt/MeOH 20:1  $\rightarrow$  5:1): *Fr. 3.1–3.15*. *Fr. 3.3* yielded **1** (11 mg), and *Fr. 3.5* was submitted to CC (*RP-18*, MeOH/ $\text{H}_2\text{O}$  1:3): **4** (9 mg) and **5** (8 mg), after further purification by CC (*Sephadex LH-20*, MeOH).

**Scrophenoside A** (=1-[4-( $\beta$ -D-Glucopyranosyloxy)-3-methoxyphenyl]ethanone 6-[4-( $\beta$ -D-Glucopyranosyloxy)-3-methoxybenzoate]; **1**): White amorphous powder (11 mg).  $[\alpha]_D^{25} = -65.1$  ( $c = 0.40$ , MeOH/H<sub>2</sub>O 1:1). IR (KBr): 3390, 2921, 1678, 1593, 1512, 1466, 1419, 1277, 1219, 1072, 1028, 763. <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 7.44 (*d*,  $J = 1.6$ , H-C(2)); 7.30 (*dd*,  $J = 1.6, 8.5$ , H-C(5)); 7.14 (*d*,  $J = 8.5$ , H-C(6)); 3.80 (*s*, MeO); 2.47 (*s*, Me(8)); 7.45 (*d*,  $J = 1.7$ , H-C(2'')); 7.18 (*d*,  $J = 8.5$ , H-C(5'')); 7.52 (*dd*,  $J = 1.7, 8.5$ , H-C(6'')); 3.77 (*s*, MeO); 5.13 (*d*,  $J = 7.2$ , H-C(1'')); 4.22 (*dd*,  $J = 7.5, 11.6$ , 1 H-C(6'')); 4.58 (*d*,  $J = 11.6$ , 1 H-C(6'')); 5.04 (*d*,  $J = 7.2$ , H-C(1''')); 3.64 (*dd*,  $J = 4.5, 12.0$ , 1 H-C(6'')); 3.45 (*br. s.*, 1 H-C(6''')). <sup>13</sup>C-NMR: Table. HR-FAB-MS: 639.1926 ( $[M - H]^-$ , C<sub>29</sub>H<sub>35</sub>O<sub>16</sub>; calc. 639.1925).

**Scrophenoside B** (=1-[4-( $\beta$ -D-Glucopyranosyloxy)phenyl]ethanone 6-[2(E)-3-(4-Hydroxyphenyl)prop-2-enoate]; **2**): White amorphous powder (15 mg).  $[\alpha]_D^{25} = -40.8$  ( $c = 0.41$ , MeOH). IR (KBr): 3326, 2927, 1693, 1659, 1610, 1582, 1516, 1449, 1282, 1257, 1066, 1045, 974, 831. <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 7.09 (*d*,  $J = 8.4$ , H-C(2), H-C(6)); 7.84 (*d*,  $J = 8.4$ , H-C(3), H-C(5)); 2.37 (*s*, Me(8)); 7.54 (*d*,  $J = 8.4$ , H-C(2'), H-C(6'')); 6.79 (*d*,  $J = 8.4$ , H-C(3'), H-C(5'')); 6.38 (*d*,  $J = 15.9$ , H-C( $\alpha$ )); 7.53 (*d*,  $J = 15.9$ , H-C( $\beta$ )); 5.06 (*d*,  $J = 7.2$ , H-C(1'')); 4.22 (*dd*,  $J = 7.5, 11.7$ , 1 H-C(6'')); 4.58 (*d*,  $J = 11.7$ , 1 H-C(6'')). <sup>13</sup>C-NMR: Table. HR-FAB-MS: 443.1355 ( $[M - H]^-$ , C<sub>23</sub>H<sub>23</sub>O<sub>9</sub>; calc. 443.1342).

**Scrophenoside C** (=4-(1-Hydroxyethyl)-2-methoxyphenyl  $\beta$ -D-Glucopyranoside; **3**): White amorphous powder (10 mg).  $[\alpha]_D^{25} = -47.8$  ( $c = 0.11$ , MeOH). IR (KBr): 3381, 2934, 1597, 1518, 1469, 1420, 1265, 1074, 1019, 808, 712. <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 6.95 (*s*, H-C(2)); 6.80 (*d*,  $J = 8.4$ , H-C(5)); 6.99 (*d*,  $J = 8.4$ , H-C(6)); 4.63 (*q*,  $J = 6.3$ , H-C(7)); 1.29 (*d*,  $J = 6.3$ , Me(8)); 3.72 (*s*, MeO); 4.82 (*d*,  $J = 7.2$ , H-C(1'')); 3.44 (*dd*,  $J = 6.0, 11.7$ , H-C(6'')); 3.64 (*d*,  $J = 11.7$ , H-C(6'')). <sup>13</sup>C-NMR: Table. HR-FAB-MS: 329.1265 ( $[M - H]^-$ , C<sub>15</sub>H<sub>21</sub>O<sub>8</sub>; calc. 329.1236).

**Scroside D** (=2-(3-Hydroxy-4-methoxyphenyl)ethyl 3-O- $\beta$ -D-Glucopyranosyl- $\beta$ -D-glucopyranoside 4-[2(E)-3-(4-Hydroxy-3-methoxyphenyl)prop-2-enoate]; **4**): Yellow amorphous powder (9 mg).  $[\alpha]_D^{25} = -44.5$  ( $c = 0.4$ , MeOH). IR (KBr): 3417, 2932, 1704, 1633, 1592, 1515, 1431, 1273, 1159, 1130, 1071, 1029, 811, 762. <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 6.62 (*s*, H-C(2)); 6.62 (*d*,  $J = 8.4$ , H-C(5)); 6.49 (*d*,  $J = 8.4$ , H-C(6)); 2.68 (*t*,  $J = 7.4$ , 2 H-C(7)); 3.88 (*m*, 1 H-C(8)); 3.60 (*m*, 1 H-C(8)); 7.30 (*s*, H-C(2'')); 6.79 (*d*,  $J = 7.7$ , H-C(5'')); 7.09 (*d*,  $J = 7.7$ , H-C(6'')); 6.47 (*d*,  $J = 15.6$ , H-C( $\alpha$ )); 7.51 (*d*,  $J = 15.6$ , H-C( $\beta$ )); 3.80 (*s*, MeO); 4.39 (*d*,  $J = 7.8$ , H-C(1'')); 3.77 (*m*, H-C(3'')); 4.69 (*t*,  $J = 9.6$ , H-C(4'')); 3.25 (*dd*,  $J = 5.4, 11.5$ , 1 H-C(6'')); 3.52 (*d*,  $J = 11.5$ , 1 H-C(6'')); 4.37 (*d*,  $J = 7.8$ , H-C(1''')). <sup>13</sup>C-NMR: Table. HR-FAB-MS: 653.2099 ( $[M - H]^-$ , C<sub>30</sub>H<sub>37</sub>O<sub>16</sub>; calc. 653.2082).

**Scroside E** (=2-(3-Hydroxy-4-methoxyphenyl)ethyl 3-O- $\beta$ -D-Glucopyranosyl- $\beta$ -D-glucopyranoside 6-[2(E)-3-(4-Hydroxy-3-methoxyphenyl)prop-2-enoate]; **5**): Yellow amorphous powder (8 mg).  $[\alpha]_D^{25} = -42.5$  ( $c = 0.40$ , MeOH). IR (KBr): 3407, 2926, 1704, 1634, 1593, 1515, 1431, 1273, 1159, 1071, 1029, 810, 762. <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 6.60 (*s*, H-C(2)); 6.80 (*d*,  $J = 8.4$ , H-C(5)); 6.43 (*d*,  $J = 8.4$ , H-C(6)); 2.66 (*t*,  $J = 7.4$ , 2 H-C(7)); 3.77 (*m*, 1 H-C(8)); 3.54 (*m*, 1 H-C(8)); 7.32 (*s*, H-C(2'')); 6.77 (*d*,  $J = 7.8$ , H-C(5'')); 7.06 (*d*,  $J = 7.8$ , H-C(6'')); 6.50 (*d*,  $J = 15.9$ , H-C( $\alpha$ )); 7.53 (*d*,  $J = 15.9$ , H-C( $\beta$ )); 3.80 (*s*, MeO); 4.37 (*d*,  $J = 7.8$ , H-C(1'')); 3.40 (*br. s.*, H-C(3'')); 3.34 (*br. d.*, H-C(4'')); 4.21 (*dd*,  $J = 6.0, 12.0$ , 1 H-C(6'')); 4.39 (*d*,  $J = 12.0$ , 1 H-C(6'')); 4.32 (*d*,  $J = 7.8$ , H-C(1''')). <sup>13</sup>C-NMR: Table. HR-FAB-MS: 653.2080 ( $[M - H]^-$ , C<sub>30</sub>H<sub>37</sub>O<sub>16</sub>; calc. 653.2082).

**Androsin** (=1-[4-( $\beta$ -D-Glucopyranosyloxy)-3-methoxyphenyl]ethanone; **6**) [6]. White amorphous solid (500 mg). <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 2.47 (*s*, COMe); 3.83 (*s*, MeO); 5.11 (*d*,  $J = 7.2$ , H-C(1'')); 7.13 (*d*,  $J = 8.4$ , H-C(6)); 7.32 (*dd*,  $J = 8.4, 1.6$ , H-C(5)); 7.45 (*d*,  $J = 1.6$ , H-C(3)). <sup>13</sup>C-NMR ((D<sub>6</sub>)DMSO, 150 MHz)<sup>1</sup>: 151.89 (C(1)); 150.03 (C(2)); 11.44 (C(3)); 131.74 (C(4)); 123.20 (C(5)); 114.82 (C(6)); 196.36 (C(7)); 26.19 (C(8)). ESI-MS (neg.): 327 ( $[M - H]^-$ ).

**Hemiphroside A** (=2-(3-Hydroxy-4-methoxyphenyl)ethyl 3-O- $\beta$ -D-Glucopyranosyl- $\beta$ -D-glucopyranoside 4-[2(E)-3-(4-Hydroxy-3-methoxyphenyl)prop-2-enoate]; **7**) [7]. Yellow amorphous powder (35 mg).  $[\alpha]_D^{25} = -46.8$  ( $c = 0.34$ , MeOH). <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 6.68 (*s*, H-C(2)); 6.81 (*d*,  $J = 8.4$ , H-C(5)); 6.63 (*d*,  $J = 8.4$ , H-C(6)); 2.72 (*t*,  $J = 7.4$ , 2 H-C(7)); 3.90 (*m*, 1 H-C(8)); 3.62 (*m*, 1 H-C(8)); 3.72 (*s*, MeO); 7.31 (*s*, H-C(2'')); 6.78 (*d*,  $J = 7.8$ , H-C(5'')); 7.10 (*d*,  $J = 7.8$ , H-C(6'')); 6.48 (*d*,  $J = 15.9$ , H-C( $\alpha$ )); 7.52 (*d*,  $J = 15.9$ , H-C( $\beta$ )); 3.81 (*s*, MeO); 4.40 (*d*,  $J = 7.8$ , H-C(1'')); 4.70 (*t*,  $J = 9.6$ , H-C(4'')); 4.37 (*d*,  $J = 7.8$ , H-C(1''')). <sup>13</sup>C-NMR: Table. ESI-MS (neg.): 667 ( $[M - H]^-$ ).

**Coniferin** (=4-[1(E)-3-Hydroxyprop-1-enyl]-2-methoxyphenyl  $\beta$ -D-Glucopyranoside; **8**) [8]. White amorphous powder (60 mg).  $[\alpha]_D^{25} = -68.3$  ( $c = 0.1$ , H<sub>2</sub>O). <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO, 600 MHz)<sup>1</sup>: 7.05 (*d*,  $J = 1.4$ , H-C(3)); 7.00 (*d*,  $J = 8.4$ , H-C(6)); 6.88 (*dd*,  $J = 8.4, 1.4$ , H-C(5)); 6.46 (*d*,  $J = 15.8$ , H-C( $\alpha$ )); 6.27 (*dt*,  $J = 15.8, 5.2$ , H-C( $\beta$ )); 4.88 (*d*,  $J = 7.2$ , H-C(1'')); 4.09 (*br.*, 2 H-C(7)). <sup>13</sup>C-NMR ((D<sub>6</sub>)DMSO, 150 MHz)<sup>1</sup>:

149.66 (C(2)); 146.63 (C(1)); 131.62 (C(4)); 129.64 (C( $\beta$ )); 129.05 (C( $\alpha$ )); 119.66 (C(5)); 115.83 (C(6)); 110.45 (C(3)); 100.6 (C(1')); 62.26 (C(7)); 56.26 (MeO). ESI-MS (neg.): 341 ( $[M-H]^-$ ).

*D-Glucopyranose 6-[(2E)-3-Phenylprop-2-enoate]* (**9**) [9]. White amorphous powder (14 mg).  $^1\text{H-NMR}$  ( $\text{CD}_3\text{OD}$ , 600 MHz): glucose moiety: 5.15 ( $d, J = 3.6$ ,  $\text{H-C}(1), \alpha\text{-D}$ ); 4.55 ( $d, J = 7.8$ ,  $\text{H-C}(1), \beta\text{-D}$ ).  $^{13}\text{C-NMR}$  ( $(\text{D}_6)\text{DMSO}$ , 150 MHz) $^1$ : cinnamoyl moiety: 167.84, 167.78 (C=O); 145.70, 145.67 (C( $\alpha$ )); 134.36 (C(1)); 130.62 (C(4)); 128.97 (C(3)); 128.14 (C(2)); 117.30, 117.26 (C( $\beta$ )); glucose moiety: 96.92 (C(1),  $\beta\text{-D}$ ); 73.49 (C(2),  $\beta\text{-D}$ ); 76.59 (C(3),  $\beta\text{-D}$ ); 70.66 (C(4),  $\beta\text{-D}$ ); 74.10 (C(5),  $\beta\text{-D}$ ); 63.95 (C(6),  $\beta\text{-D}$ ); 92.76 (C(1),  $\alpha\text{-D}$ ); 74.91 (C(2),  $\alpha\text{-D}$ ); 72.43 (C(3),  $\alpha\text{-D}$ ); 69.56 (C(4),  $\alpha\text{-D}$ ); 70.46 (C(5),  $\alpha\text{-D}$ ); 63.91 (C(6),  $\alpha\text{-D}$ ). ESI-MS (neg.): 309 ( $[M-H]^-$ ).

*D-Glucopyranose 6-[(2E)-3-(4-Hydroxyphenyl)prop-2-enoate]* (**10**) [10]. White amorphous powder (12 mg).  $^1\text{H-NMR}$  ( $(\text{D}_6)\text{DMSO}$ , 600 MHz) $^1$ : glucose moiety: 4.91 ( $d, J = 4.1$ ,  $\text{H-C}(1), \alpha\text{-D}$ ); 4.31 ( $d, J = 7.3$ ,  $\text{H-C}(1), \beta\text{-D}$ ); *p*-coumaroyl moiety: 6.78 ( $d, J = 8.6$ ,  $\text{H-C}(2), \text{H-C}(6)$ ); 7.54 ( $d, J = 8.6$ ,  $\text{H-C}(3), \text{H-C}(5)$ ); 6.39 ( $d, J = 15.8$ ,  $\text{H-C}(\alpha)$ ); 7.53 ( $d, J = 15.8$ ,  $\text{H-C}(\beta)$ ).  $^{13}\text{C-NMR}$  ( $(\text{D}_6)\text{DMSO}$ , 150 MHz) $^1$ : *p*-coumaroyl moiety: 167.35 (C=O); 145.62, 145.49 (C( $\alpha$ )); 131.05 (C(2)); 125.72 (C(1)); 116.43 (C(3)); 114.73, 114.62 (C( $\beta$ )); glucose moiety: 97.58 (C(1),  $\beta\text{-D}$ ); 73.56 (C(2),  $\beta\text{-D}$ ); 77.09 (C(3),  $\beta\text{-D}$ ); 70.88 (C(4),  $\beta\text{-D}$ ); 75.37 (C(5),  $\beta\text{-D}$ ); 92.97 (C(1),  $\alpha\text{-D}$ ); 74.25 (C(2),  $\alpha\text{-D}$ ); 72.85 (C(3),  $\alpha\text{-D}$ ); 69.56 (C(4),  $\alpha\text{-D}$ ); 71.31 (C(5),  $\alpha\text{-D}$ ); 64.69, 64.63 (C(6),  $\alpha\text{-D}$  and  $\beta\text{-D}$ ). ESI-MS (neg.): 325 ( $[M-H]^-$ ).

*TLC Analysis of Sugars of 1–5*. Compounds **1–5** were each applied to a TLC plate and then hydrolyzed under HCl vapor at 60° for 40 min. After the excess HCl was removed, glucose was applied to the same plate, the plate developed ( $\text{CHCl}_3/\text{MeOH}/\text{H}_2\text{O}/\text{AcOH}$  16:9:2:2), sprayed with aniline/phthalic acid, and heated. Glucose gave a purple spot,  $R_f$  0.42.

## REFERENCES

- [1] H. Zhang, Z. Zhang, 'Handbook of Chinese Traditional Medicine Resources', Science Press, Beijing, 1994, p. 1149.
- [2] P. S. Xie, *ZhongCaoYao* **1983**, 14, 341.
- [3] H. F. Smit, A. J. J. Van den Berg, B. H. Kroes, C. J. Beukelman, H. C. Quarles van Ufford, H. van Dijk, R. P. Labadie, *J. Nat. Prod.* **2000**, 63, 1300.
- [4] D. Q. Wang, Z. D. He, B. S. Feng, C. R. Yang, *Acta Botanica Yunnanica* **1993**, 15, 83.
- [5] J. X. Li, P. Li, Y. Tezwca, T. Namba, S. Kadota, *Phytochemistry* **1998**, 48, 537.
- [6] H. Stuppner, H. Wagner, *Planta Med.* **1989**, 55, 467.
- [7] W. G. Ma, X. C. Li, Y. U. Qing, Q. S. Li, C. R. Yang, *Acta Botanica Yunnanica* **1995**, 17, 96.
- [8] X. L. Shen, Y. M. Shen, Y. J. Hu, Q. Z. Mu, *ZhongCaoYao* **1996**, 27, 259.
- [9] H. Shimamura, Y. Sashida, T. Adachi, *Phytochemistry* **1988**, 27, 641.
- [10] Y. Kashiwada, G. Nonaka, I. Nishioka, T. Yamagishi, *Phytochemistry* **1988**, 27, 1473.
- [11] W. Dorsch, H. Stuppner, H. Wagner, M. Gropp, S. Demoulin, J. Ring, *Int. Arch. Allergy Appl. Immunol.* **1991**, 95, 128.

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